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Phil. Trans. R. Soc. Lond. B 1997 **352**, 1327-1330 doi: 10.1098/rstb.1997.0117

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The importance of the back-signal from T cells into antigen-presenting cells in determining susceptibility to parasites

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SUMMARY

It has long been known that certain MHC class II genes can dominantly suppress immune responses and so increase susceptibility to parasite infections, but the mechanism has been unclear. Recent work has revealed one way in which this form of suppression may operate, through gating by MHC class II molecules of the back-signal from activated T cells into macrophages. The two known suppressive genes of the mouse are expressed in macrophages more extensively than are other class II genes. This is assocciated with suppression of IL-4 production resulting, we infer, from overproduction in the macrophages of IL-12, the counter-cytokine to IL-4. The lack of IL-4 may itself be immunosuppressive, even for Th2 responses, and excess IL-12 can overinduce the antiproliferative cytokine IFN-gamma. Although this mechanism requires further substantiation, we believe that it offers a reasonable answer to an old conundrum.

The ability of certain class II MHC genes to suppress antiparasite immune responses began to attract attention during the 1980s. Expression in the mouse of H2E was found to reduce the protective response to nematode infection (Wassom et al. 1987) and to visceral leishmaniasis (Blackwell & Roberts 1987), although the effect is not always obtained (Behnke & Wahid 1991). In man, expression of certain HLA.DQ alleles reduced the response to schistosome antigens (Hirayama et al. 1987). It has been suggested that genes responsible for this type of activity might be retained in natural populations by the selective advantage conferred by reducing damage from inflammation (Mitchison & Oliveira 1986). However, little progress was made, mainly because the mechanism of suppression was not understood. That problem has now been solved, in part at least, as summarized here. The solution turns on polymorphism in the promoters of the class II MHC genes in question.

We were led to investigate this type of polymorphism by our longstanding interest in the protective and suppressive function of MHC class II genes (Silver & Lane 1975). The first sign of an effect unlikely to be mediated by the classical exon function of determinant selection was the discovery that expression of the H2Egene suppressed the immune response not only to foreign lactic dehydrogenase (Nagy *et al.* 1981), but also to many other antigens (Oliveira & Mitchison 1989). Particularly relevant was the finding that the response to a whole range of structurally linked antigens could be suppressed at the same time, such as that to an assembly of non-MHC alloantigens presented on the same cell (Krzych *et al.* 1989); much the same must have occurred in the suppression of the antiparasite responses mentioned above. Subsequently the *b* allele at H2A was found to have a similar but weaker effect, which operates additively with that of H2E (Hesse *et al.* 1996). This line of research has now moved on to the prevention and cure of autoimmune disease in mouse models (Hirose *et al.* 1994; Gonzalez-Gay *et al.* 1994; Mitchison & Brunner 1995). It has obvious relevance to human immunological diseases, where this genetic approach to cure makes a welcome change from the dauntingly difficult task of determining the causation of these diseases (Mitchison 1992).

'Suppression' is an appropriate term to use, as these genes are effective in a single dose and must therefore play an active role, rather than one of passive inactivation. Their activity in this respect represents no more than a bias, as they can also function as normal positive immune response genes. There is evidence of similar effects in man, particularly among DQ and DR2, 5 and 7 haplotypes; and an allele that is otherwise associated with protection may cease to be protective in a disease for which it is a susceptibility factor, as in the case of juvenile chronic arthritis and perhaps also in multiple sclerosis (Guardiola *et al.* 1996).

Some MHC class II promoters are highly polymorphic, to an extent that indicates that they must be under selective pressure (Guardiola *et al.* 1996). In contrast, MHC class I promoters are relatively constant (Cereb & Yang 1994; Yao *et al.* 1995). Especially telling is the finding that sequence variation

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between allelic promoters in the mouse is actually greater within (and immediately adjacent to) the X and Y boxes than in the intervening sequences (Janitz et al. 1997). These boxes are where transcription factors are known to bind, and where sequence is conserved over long evolutionary distance (Benoist & Mathis 1990; Sultmann et al. 1993); indeed this conservation was one of their original defining features. The situation is slightly different in man, where more alleles have been sequenced, and variation within the X and Y boxes was again found to be high, but not quite as much so as in the intervening sequences (Louis et al. 1993). For both man and mouse these generalizations are provisional, as there are many more promoters to sequence; it would be of interest to learn about promoter diversity within the mouse genus, for example. What seems clear is that the functionally active part of these promoters must be under selective pressure to diversify, presumably reflecting increased fitness of heterozygotes.

Analysis of cytokine gene expression provides a clue to the mode of action of these suppressive/protective genes. A survey of the major T-cell cytokines revealed that the most conspicuous effect of the $H2A^b$ allele is to inhibit the burst of IL-4 transcription that occurs soon after immunization in a suppressible immune response; *in vivo* treatment with anti-IL-4 monoclonal antibody mimics this effect, provided that the antibody is given immediately upon immunization (Brunner *et al.* 1995; Hesse *et al.* 1996). As the suppressive allele is known to play a positive role, a reasonable inference is that it elicits a counter-cytokine to IL-4.

Our most recent finding is that the suppressive genes $H2A^b$ and H2E are both expressed more extensively in activated macrophages than are the neutral alleles $H2A^d$, $H2A^k$ and $H2A^q$. This parallels an earlier finding that promoter–reporter gene constructs transfected into a macrophage cell line yield a higher signal when made with the $H2A^b$ promoter. The increase results from a single $A \rightarrow G$ substitution at the 3' end of the X box in the $H2A^b$ promoter, which is reversible by site-specific mutagenesis (Janitz *et al.* 1997).

Recent discoveries concerning IL-12 offer a solution to this puzzle, shown in figure 1. IL-12 is the principal counter-cytokine to IL-4 (Seder et al. 1996; Abbas et al. 1996). It is made mainly by activated macrophages and dendritic cells, in response to back-signals delivered by recently activated T cells. The ligand-pair that actually delivers the signal is CD40L on the T cell ligating to CD40 on the antigen-presenting cell (APC), but signalling is facilitated by ligation of MHC class II molecules on the APC to the T-cell receptor (Kato et al. 1995; Koch et al. 1996; Cella et al. 1996). It is reasonable to suppose that the level of expression of MHC class II gates the strength of this signal, so that increased expression leads to greater production of IL-12, which would in turn inhibit IL-4 production. One may also suppose, as shown in figure 1, that these interactions take place among T cells clustering around an APC (although not necessarily all at the same time). The IL-12-inducing T cell binds via a suppressive MHC class II molecule, and the target Tcell, which would otherwise mediate a positive response, binds via a positively-inducing class II

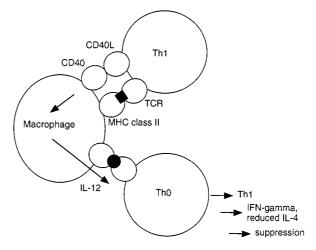


Figure 1. Epicrine interaction between a T cell recognizing a suppressive epitope (solid square) and one recognizing what would otherwise be a positive-response-inducing epitope (solid circle). The level of expression of the MHC class II molecule recognizing the suppressive epitope gates the back signal (delivered via CD 40L) into the macrophage; heightened expression causes higher IL-12 production. The IL-12 is envisaged as having a suppressive effect via IFN-gamma induction (Caspi *et al.* 1997) and reduced IL-4 levels.

molecule. An interaction of this sort between two T cells, which occurs via a change induced in their common APC, has been termed 'epicrine' (T. Tada, personal communication), a useful term.

The only surprising feature of this hypothesis is the suppressive role of IL-12, a molecule which on the basis of in vitro experiments is considered essential for development of a Thl response (Abbas et al. 1996). Collagen-induced arthritis, known to be susceptible to the suppressive effect of $H2A^b$ and H2E, is mainly a Thl disease like other organ-specific autoimmunities induced with complete Freund's adjuvant. However, there is something odd about these diseases. Although IL-12 given during the course of the disease has an exacerbating effect, when given at the time of induction it is protective (Hess et al. 1996; Caspi et al. 1997). This mirrors the timing seen with anti-IL-4 treatment referred to above. The protective effect appears to operate via induction of IFN-gamma, because knockout of this gene exacerbates at least one of these diseases (Krakowski & Owens 1996) and blocks the protective effect of early treatment with IL-12 in another (Caspi et al. 1997). All in all, the picture derived from in vitro studies on TCR-transgenic T cells (Abbas et al. 1996) may be oversimplified. It seems likely that an important distinction applies in vivo between an initial proliferative phase of the immune response, in which IL-4 may be important as a growth factor for both Thl and Th2 cells and IFN-gamma as an antiproliferative agent (Chiodetti & Schwartz 1995; Krakowski & Owens 1996), and a later phase in which the role of these cytokines in maintaining Thl/Th2 balance becomes more important; the role of IL-12 in the second phase is the subject of a recent review (Seder et al. 1996).

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The scheme presented in figure l is unlikely to be the only mechanism by which differential expression of MHC class II genes influences the immune response. Suppressive MHC molecules have hardly begun to be studied in man, where the lack of suitable monoclonal antibodies is an obstacle. What is presented here is simply the first mechanism to be worked out in detail. It will no doubt be tested in future investigations, but there are many other possible mechanisms (Gonzalez-Gay *et al.* 1994; Guardiola *et al.* 1997).

To complete this presentation, a note about the association of promoter and exon variation. An extreme position would be that the polymorphism observed in promoters merely reflects-is dragged along by-the well-known polymorphism of exons. This seems increasingly unlikely in view of the arguments deployed above. However, it is unlikely that the two are entirely independent of one another. Very little is known about why particular MHC alleles are retained in natural populations, although it is generally assumed that this reflects the need to present the antigens of pathogens. Suppression of the response to pathogen antigens would also be appropriate under certain circumstances, particularly where chronic immunopathology would otherwise develop (Mitchison & Oliveira 1986); leprosy is a case in point, where an immunosuppressive, dominant, HLA-linked effect favours progress into the less life-threatening lepromatous form of the disease (van Eden et al. 1985), a development which is beneficial to both host and pathogen. In a scenario of this sort it is not difficult to imagine how a suppressive level of expression could come to associate, during the course of evolution, with the ability to present a particular set of epitopes.

The argument presented here is that an early burst of IL-12 production can suppress the immune response independent of (or at least in addition to) itsThl-inducing effect. If so, it clearly presents a way of subverting the immune response that is likely to have attracted the attention of parasites. It is true that mice respond to the injection of large doses of Leishmania major in the opposite way, by producing an early burst of IL-4 (Launois et al. 1995). In the light of the Blackwell & Roberts (1987) findings, the hypothesis advanced here makes the following sharp prediction: when administered by the route and in the dose used in the that earlier study, L. donovani should elicit less early IL-4 and more early IL-12 in mice that express H2E and $H2A^{b}$ than in control, non-expressing mice of the same genetic background (e.g. in Bl0.A(5R) versus Bl0.A(4R) or Bl0.Q). In summary, the possibility that parasite-mediated suppression may on occasion be detrimental to the host should not be neglected, and we should be alert to the danger that precocious induction of IL-12 may present.

REFERENCES

- Abbas, A. K., Murphy, K. M. & Sher, A. 1996 Functional diversity of helper T lymphocytes. *Nature* 383, 787–793.
- Behnke, J. M. & Wahid, F. N. 1991 Immunological relationships during primary infection with Heligmosomoides polygyrus (*Nematospiroides dubius*): H-2 linked genes determine worm survival. *Parasitology* 1, 157–164.

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- Blackwell, J. M. & Roberts, M. B. 1987 Immunomodulation of murine visceral leishmaniasis by administration of monoclonal anti-Ia antibodies: differential effects of anti-I-A vs. anti-I-E antibodies. *Eur. J. Immunol.* 17, 1669–1672.
- Benoist, C. & Mathis, D. 1990 Regulation of major histocompatibility complex class II genes: X and Y and other letters of the alphabet. A. Rev. Immunol.8, 681–715.
- Brunner, M. C., Larsen, S., Sette, A. & Mitchison, N. A. 1995 Altered Thl/Th2 balance associated with the immunosuppressive/protective effect of the H-2Ab allele on the response to allo-HPPD. *Eur. J. Immunol.* 25, 3285–3289.
- Caspi, R. R., Silver, P. B., Agarwal, R., Rizzo, L. V., Chan, C. C. & Tarrant, T. 1997 Systemic administration of IL-12 protects mice from experimental autoimmune uveitis (EAU) through a mechanism involving IFN-gamma. *J* . *Allergy. Clin. Immunol.* (In the press.)
- Cella, M., Scheidegger, D., Palmer Lehmann, K., Lane, P., Lanzavecchia, A. & Alber, G. 1996 Ligation of CD40 on dendritic cells triggers production of high levels of interleukin-12 and enhances T cell stimulatory capacity: T-T help via APC activation. *J. Exp. Med.* 184, 747–752.
- Cereb, N. & Yang, S. Y. 1994 The regulatory complex of HLA class I promoters exhibits locus-specific conservation with limited allelic variation. *J. Immunol.* **152**, 3873–3883.
- Chiodetti, N. & Schwartz, R. H. 1995 The role of CD28 in the activation of T lymphocytes to proliferate in response to IL4. *Res. Immunol.* **146**, 169–171.
- Gonzalez-Gay, M. A., Nabozny, G. H., Bull, M. J., Zanelli, E., Douhan, J., Griffiths, M. M., Glimcher, L. H., Luthra, H. S. & David, C. S. 1994 Protective role of major histocompatibility complex class II Ebd transgene on collageninduced arthritis. *J. Exp. Med.* 180, 1559–1564.
- Guardiola, J., Maffei, A., Lauster, R., Mitchison, N. A., Accolla, R. S. & Sartoris, S. 1996 Functional significance of polymorphism among MHC class II gene promoters. *Tissue Antigens* 48, 615–625.
- Hess, H., Gately, M. K., Rude, E., Schmitt, E., Szeliga, J. & Germann, T. 1996 High doses of interleukin-12 inhibit the development of joint disease in DBA/1 mice immunized with type II collagen in complete Freund's adjuvant. *Eur. J. Immunol.* 26, 187–191.
- Hesse, M., Bayrak, S. & Mitchison, A. 1996 Protective major histocompatibility complex genes and the role of interleukin-4 in collagen-induced arthritis. *Eur. J. Immunol.* 26, 3234–3237.
- Hirayama, K., Matsushita, S., Kikuchi, I., Iuchi, M., Ohta, N., & Sasazuki, T. 1987 HLA-DQ is epistatic to HLA-DR in controlling the immune response to schistosomal antigen in humans. *Nature* **327**, 426–430.
- Hirose, S., Zhang, D., Nozawa, S., Nishimura, H. & Shirai, T. 1994 The E-linked subregion of the major histocompatibility complex down-regulates autoimmunity in NZB x NZW F1 mice. *Immunogenetics* **40**, 150–153.
- Janitz, M., Mitchison, A., Reiners-Schramm, L. & Lauster, R. 1997 Polymorphic MHC class II promoters exhibit distinct patterns in various antigen presenting cell lines. *Tissue Antigens* 49, 99–106.
- Kato, T., Hakamada, R., Yamane, H. & Nariuchi, H. 1996 Induction of IL-12 p40 messenger RNA expression and IL-12 production of macrophages via CD40-CD40 ligand interaction. *J. Immunol.* **156**, 3932–3938.
- Koch, F., Stanzl, U., Jennewein, P., Janke, K., Heufler, C., Kampgen, E., Romani, N. & Schuler, G. 1996 High level IL-12 production by murine dendritic cells: upregulation via MHC class II and CD40 molecules and downregulation by IL-4 and IL-10. *J. Exp. Med.* **184**, 741–746.
- Krakowski, M. & Owens, T. 1996 Interferon-gamma confers resistance to experimental allergic encephalomyelitis. *Eur. J. Immunol.* 26, 1641–1646.
- Krzych, U., Nanda, N. & Sercarz, E. 1989 Specificity and interactions of CD8+ T suppressor cells. *Res. Immunol.* 140, 302–307.

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- Launois, P., Ohteki, T., Swihart, K., MacDonald, H. R., & Louis, J. A. 1995 In susceptible mice, *Leishmania major* induces very rapid interleukin-4 production by CD4⁺ T cells which are NK1.1⁺. *Eur. J. Immunol.* 25, 3298–3307.
- Louis, P., Eliaou, J. F., Kerlan, C. S., Pinet, V., Vincent, R. & Clot, J. 1993 Polymorphism in the regulatory region of HLA-DRB genes correlating with haplotype evolution. *Immunogenetics* 38, 21–26.
- Mitchison, N. A. 1992 Specialization, tolerance, memory, competition, latency, and strife among T cells. A. Rev. Immunol. 10, 1–12.
- Mitchison, N. A. & Brunner, M. C. 1995 Association of H2Ab with resistance to collagen-induced arthritis in H2-recombinant mouse strains: an allele associated with reduction of several apparently unrelated responses. *Immunogenetics* 41, 239–245.
- Mitchison, N. A. & Oliveira, D. B. G. 1986 Chronic infection as a major force in the evolution of the suppressor T cell system. *Parasitol. Today* **2**, 312–313.
- Nagy, Z. A., Baxevanis, C. N., Ishii, N. & Klein, J. 1981 Ia antigens as restriction molecules in Ir-gene controlled Tcell proliferation. *Immunol. Rev.* 60, 59–83.

- Oliveira, D. B. & Mitchison, N. A. 1989 Immune suppression genes. *Clin. Exp. Immunol.* **75**, 167–177.
- Seder, R. A., Kelsoll, B. L. & Jankovic, D. 1996 Differential roles of IL-12 in the maintenance of immune responses to infections versus autoimmune disease. *J. Immunol.* 157, 2745–2748.
- Silver, D. M. & Lane, D. P. 1975 Dominant nonresponsiveness in the induction of autoimmunity to liver-specific Fantigen. *J. Exp. Med.* 142, 1455–1461.
- Sultmann, H., Mayer, W. E., Figueroa, F., O'hUigin, C. & Klein, J. 1993 Zebrafish Mhc class II alpha chain-encoding genes: polymorphism, expression, and function. *Immunogenetics* 38, 408–420.
- van Eden, W., Gonzalez, N. M., de Vries, R. R., Convit, J. & van Rood, J. J. 1985 HLA-linked control of predisposition to lepromatous leprosy. *J. Infect. Dis.* 151, 9–14.
- Wassom, D. L., Krco, C. J. & David, C. S. 1987 I-E expression and susceptibility to parasite infection. *Immunol. Today* 8, 39–43.
- Yao, Z., Volgger, A., Scholz, S. & Albert, E. D. 1995 Sequence polymorphism in the HLA-B promoter region. *Immunogenetics* 41, 343–353.